

XXXI Symposium on Bioinformatics and Computer-Aided Drug Discovery

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Protein Engineering Methods for Challenging Membrane-Bound Drug Targets

Ivan Gushchin

October 22nd, 2025

Molecular interactions at the basis of disease



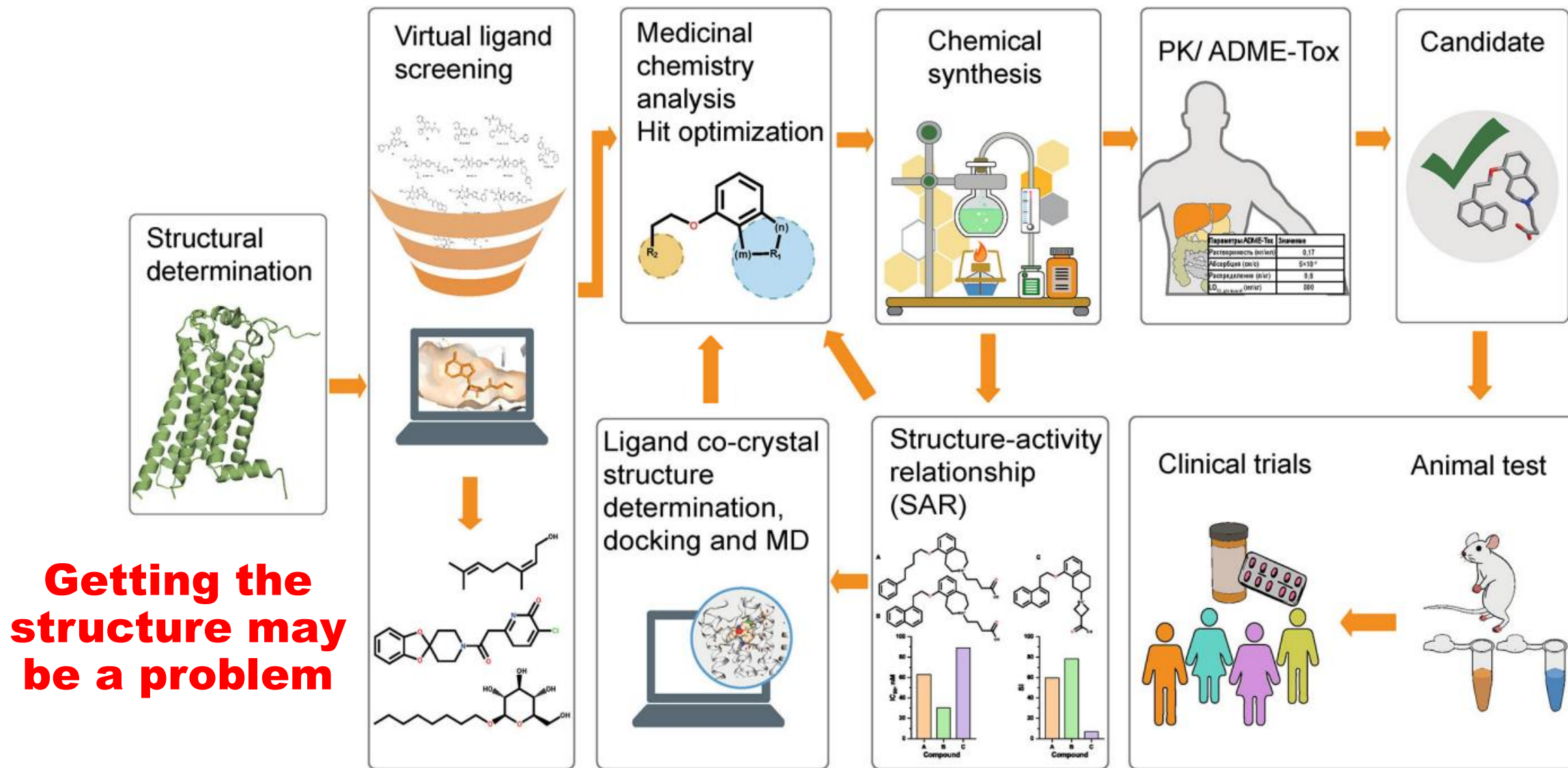
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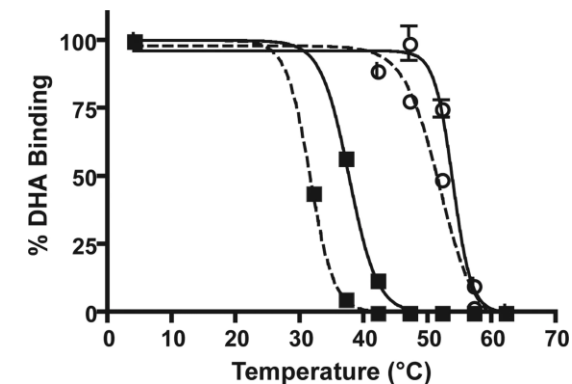
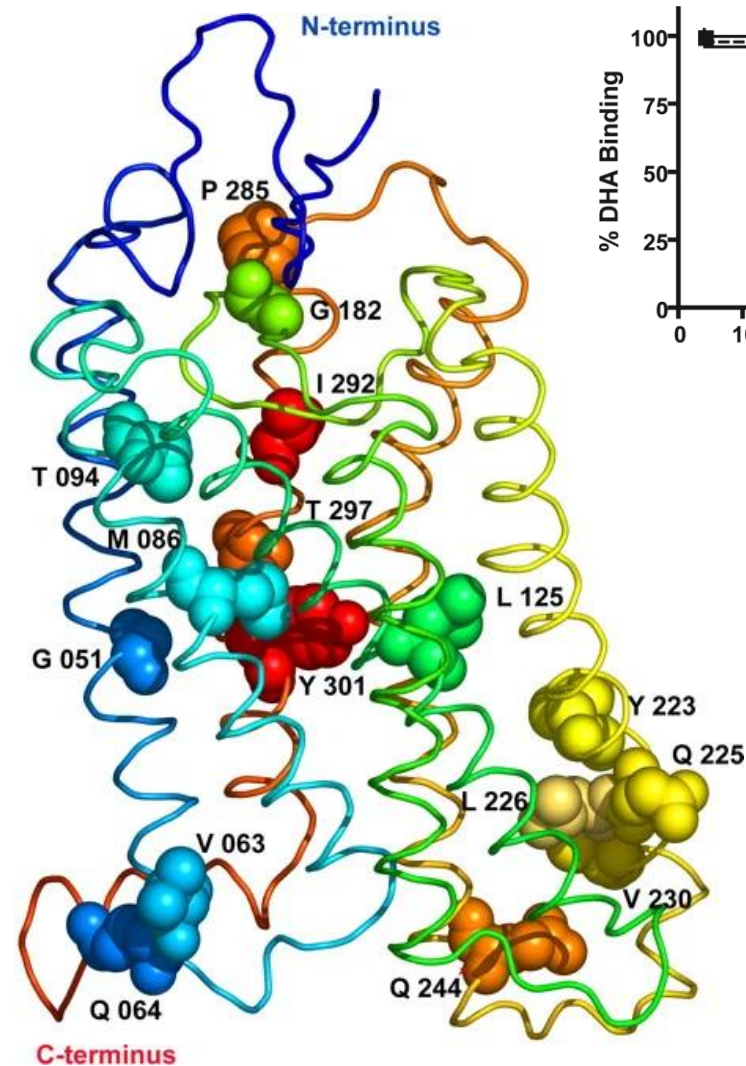
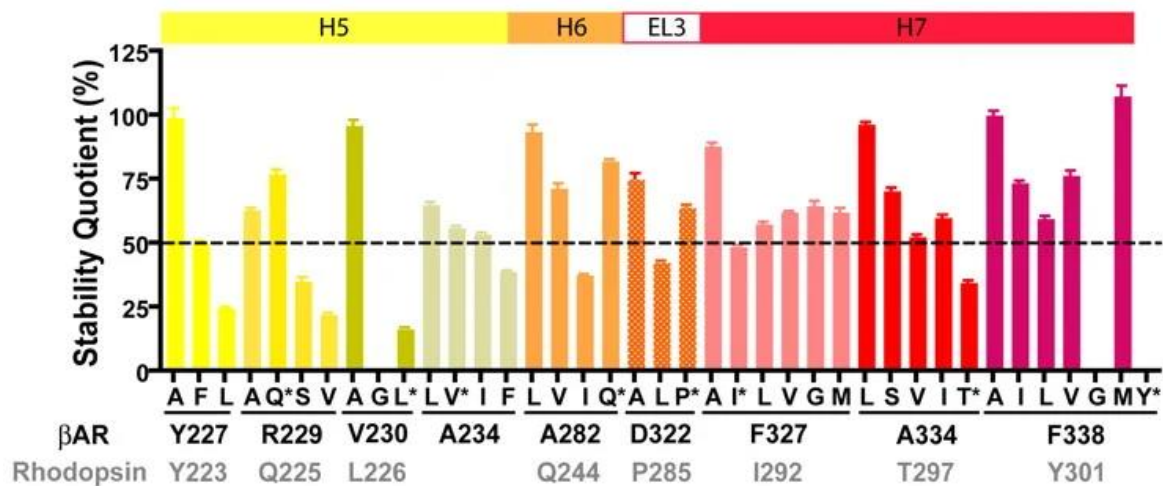
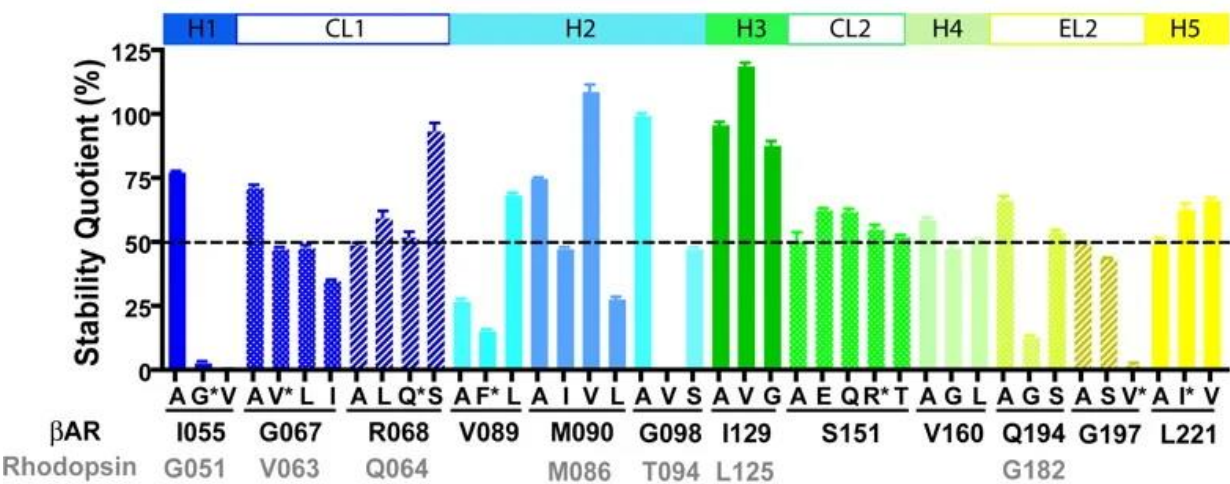
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Need to identify the target & disrupt (or enable) interactions

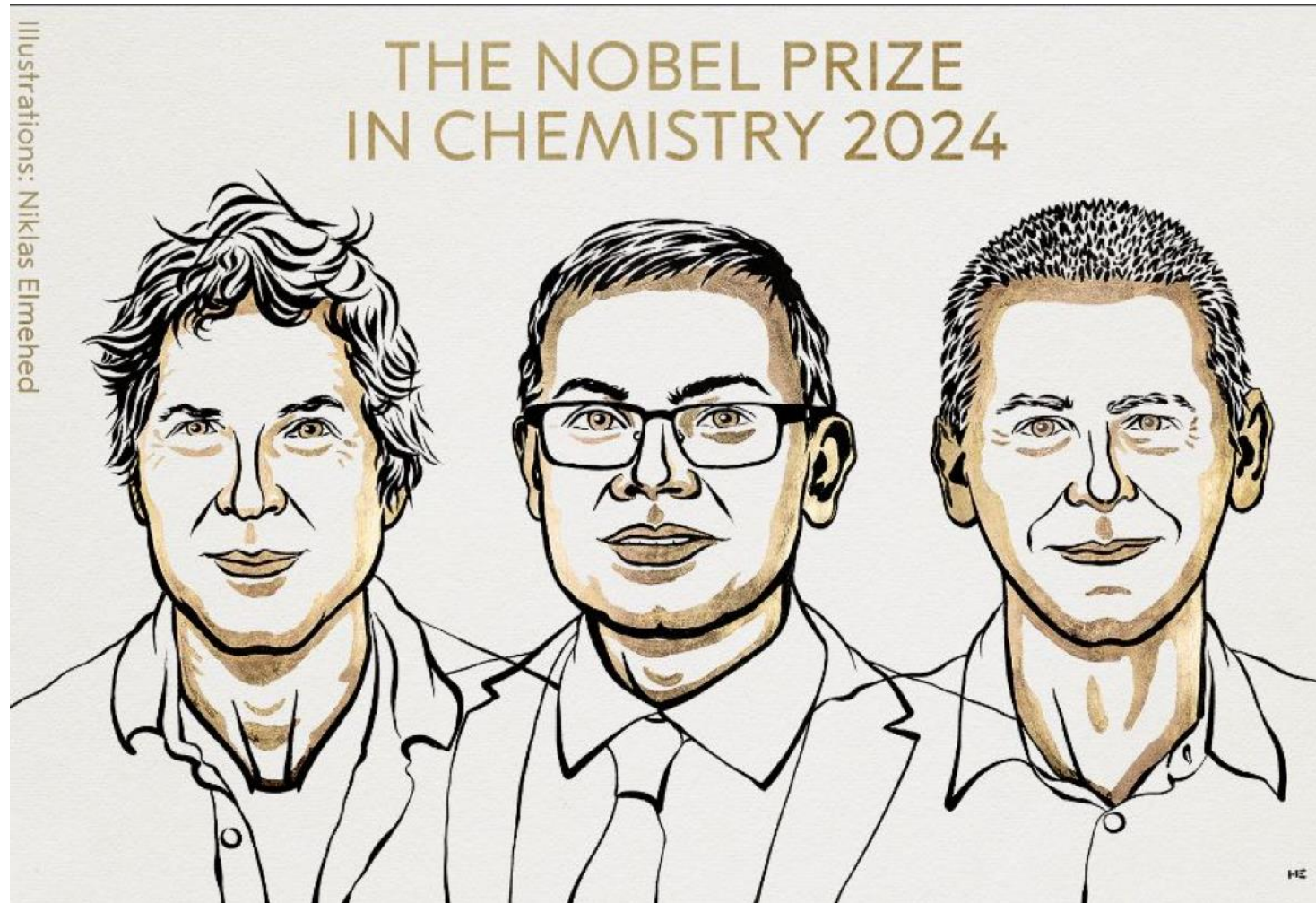
Structure-based drug development



Protein stabilization before AI



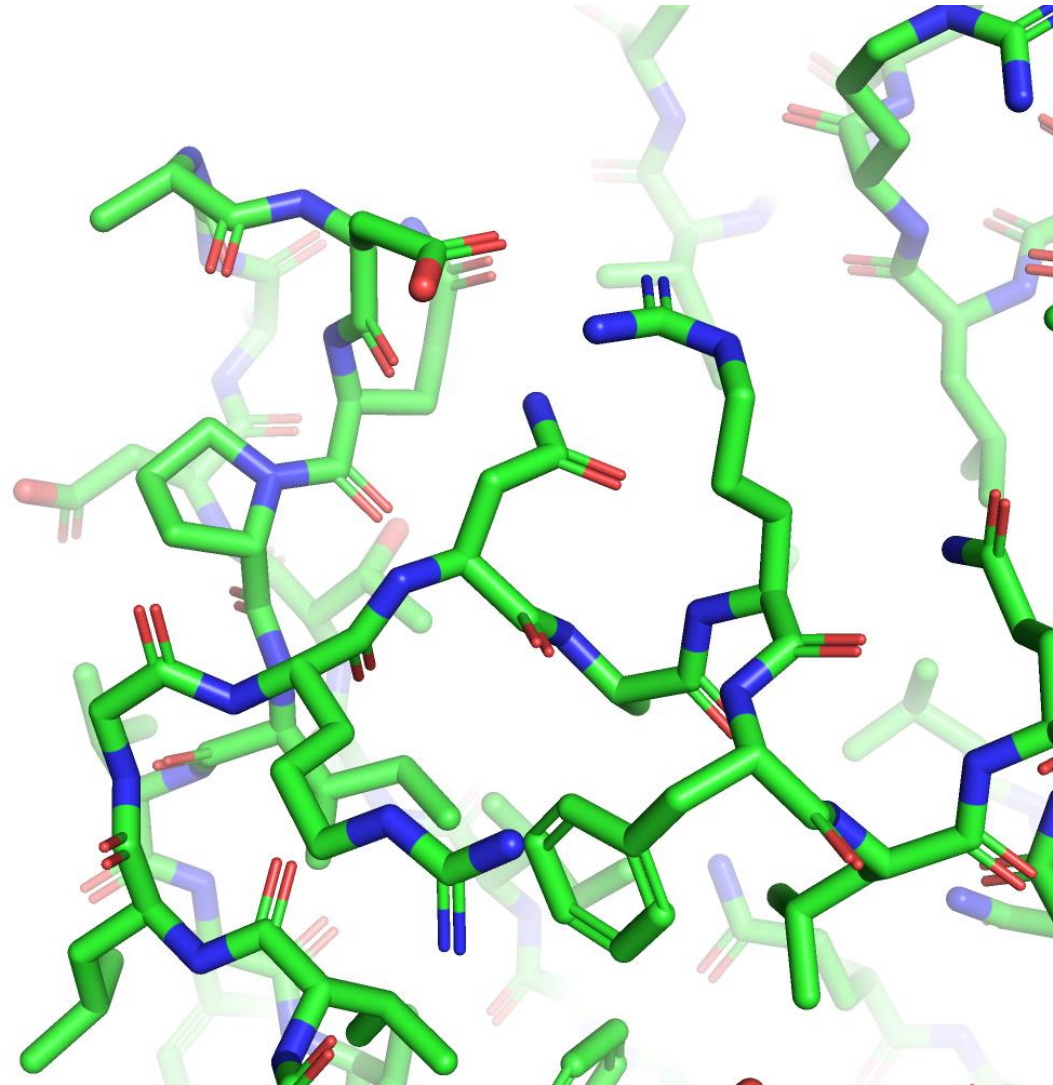
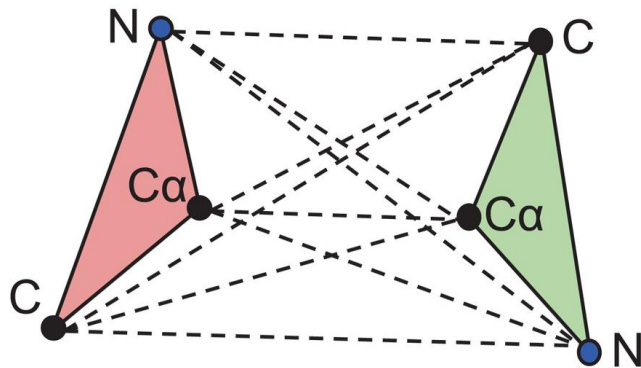
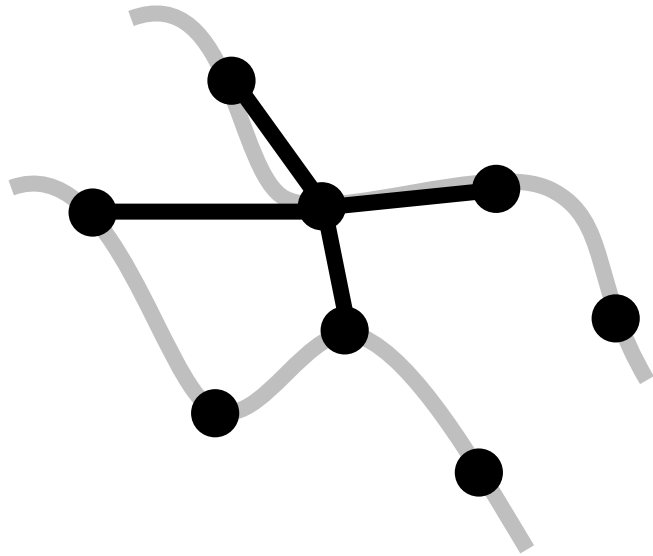
AI triumphs in structural biology



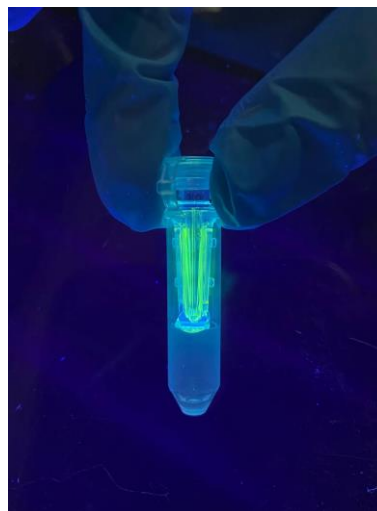
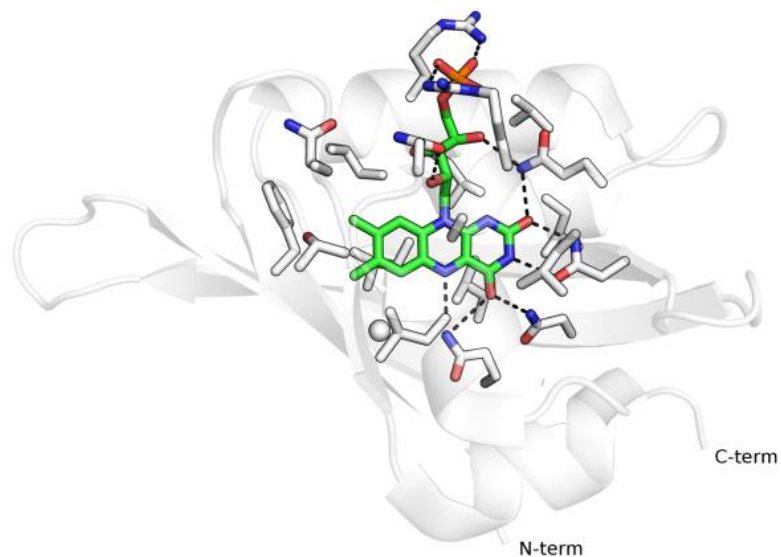
“for protein structure prediction”

“for computational protein design”

ProteinMPNN: identification of “optimal” amino acids



Protein stabilization after AI

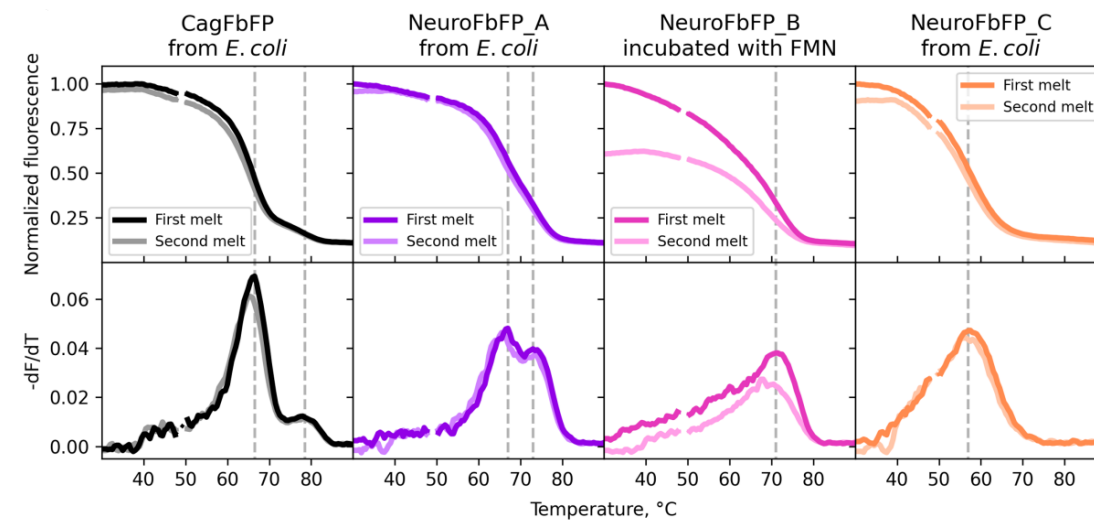
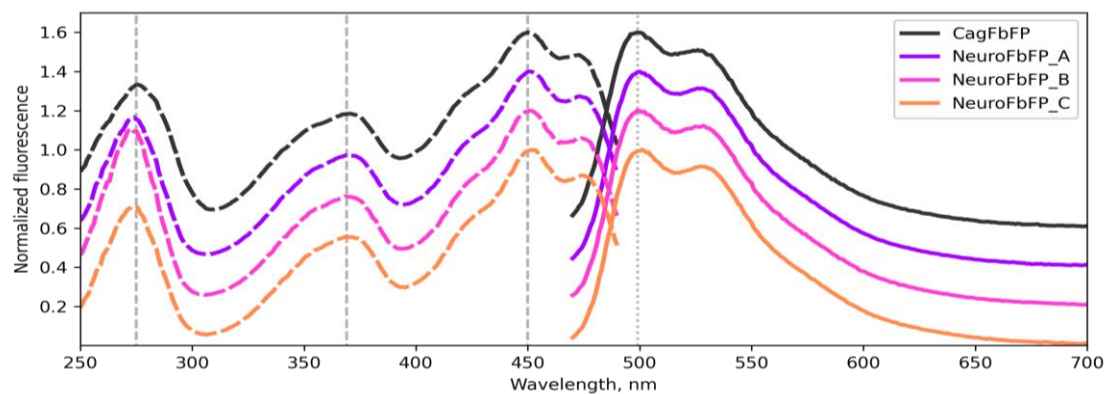


55-66% sequence identity

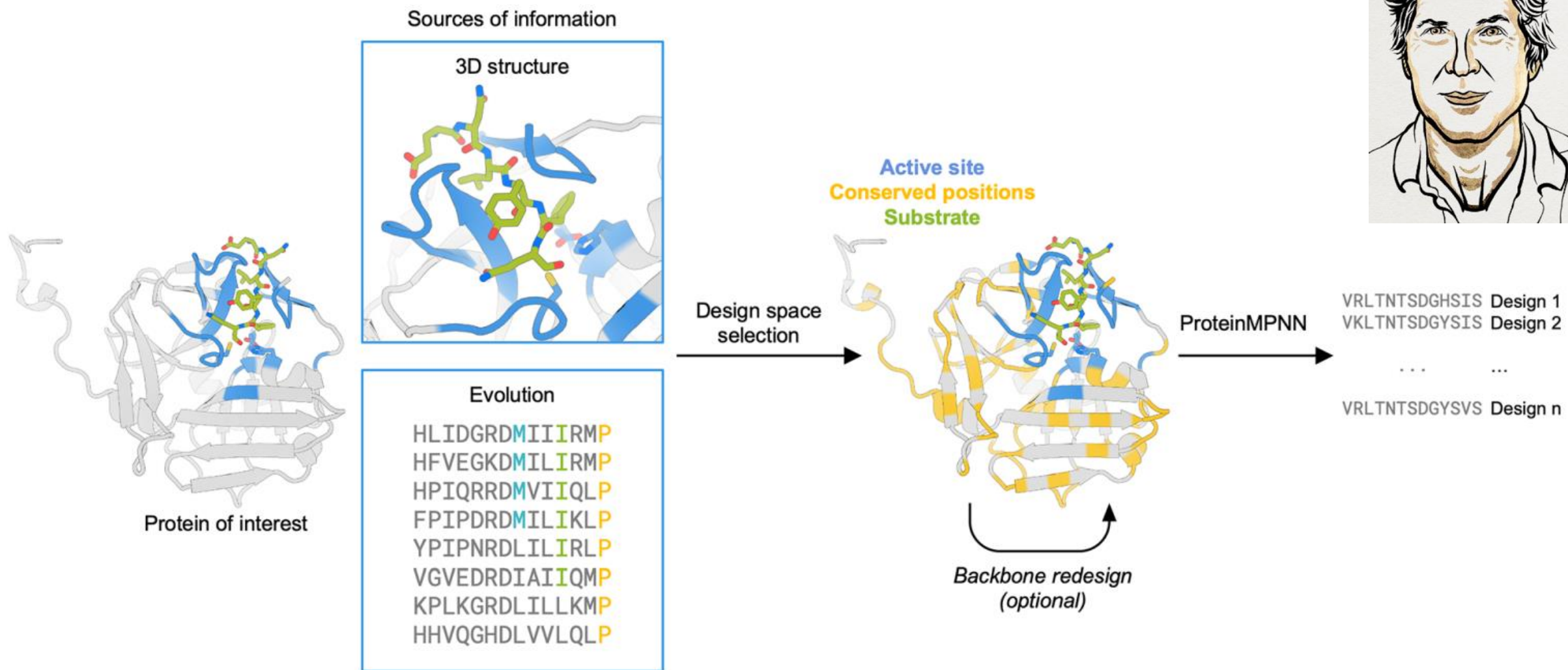


Original

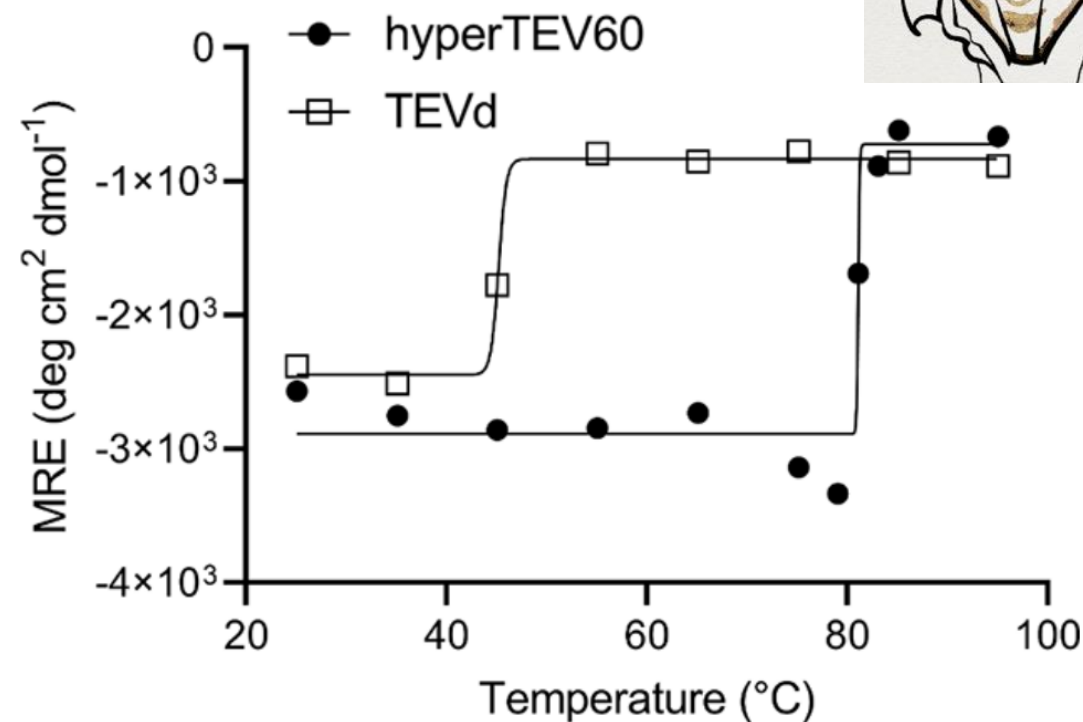
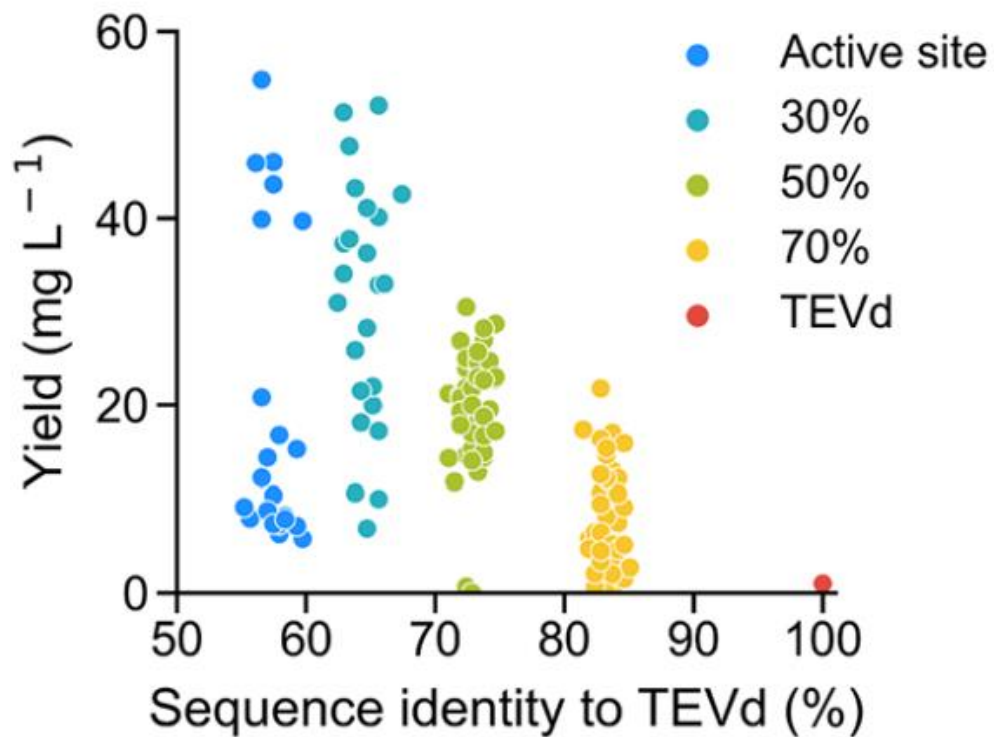
Engineered



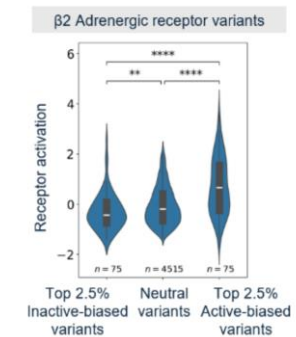
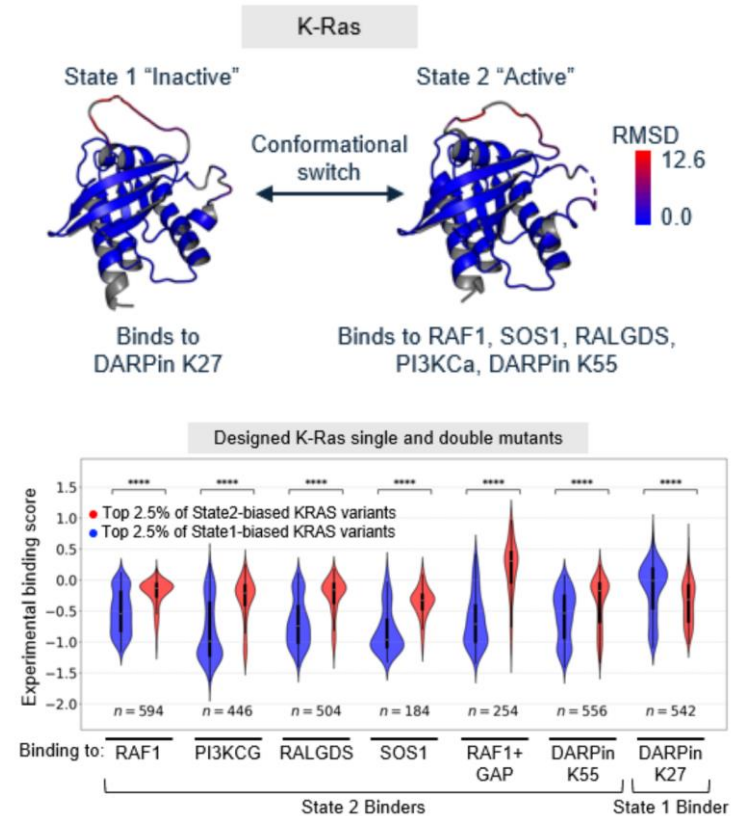
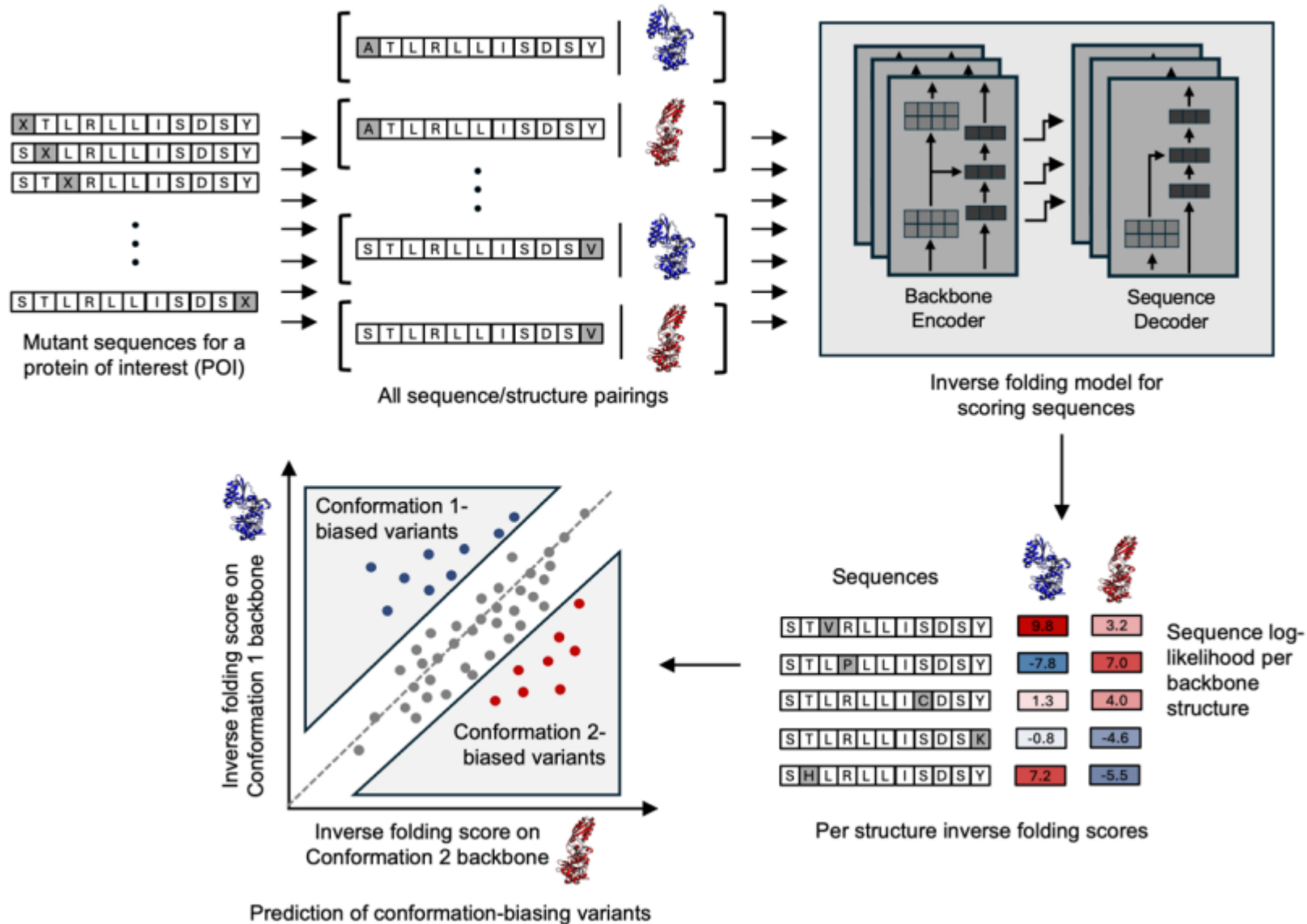
Protein stabilization after AI



Protein stabilization after AI



Stabilization of a specific conformation

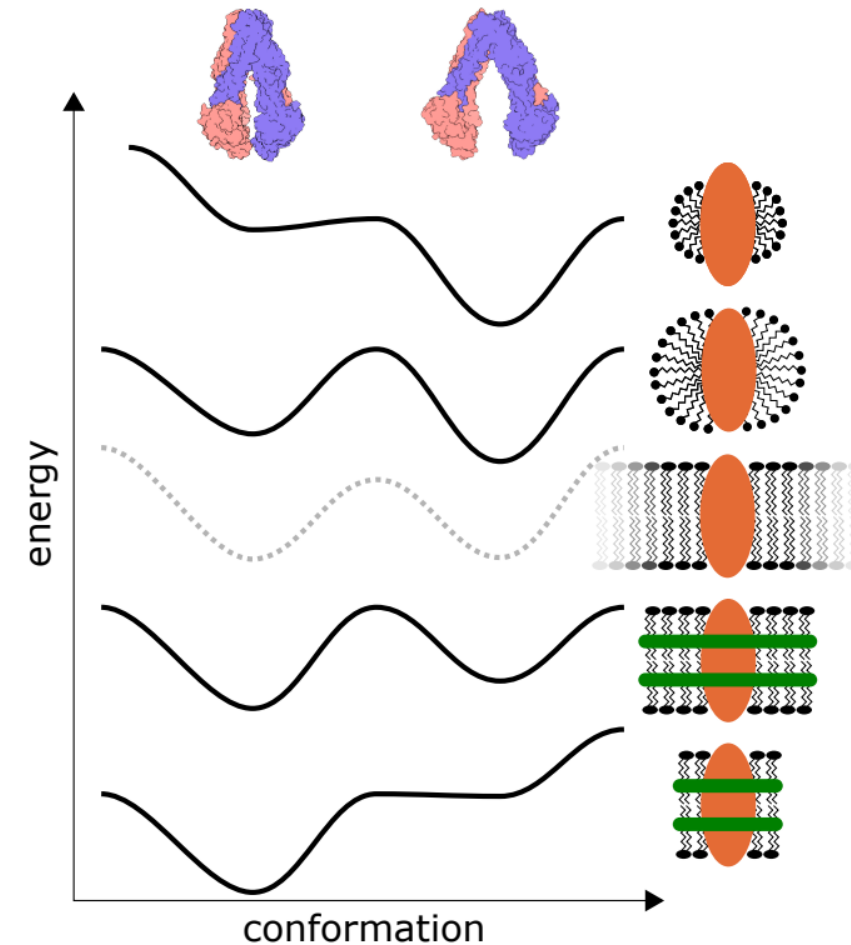
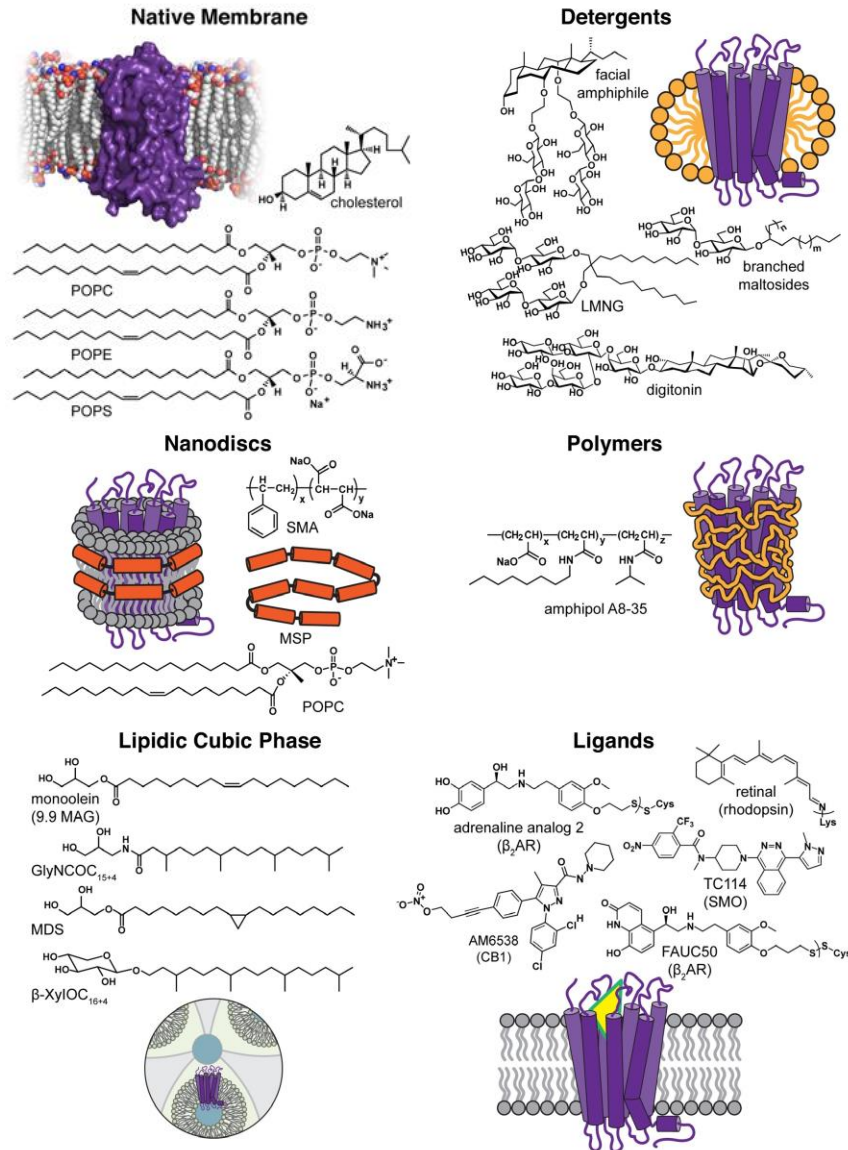


<https://doi.org/10.7554/eLife.54895>

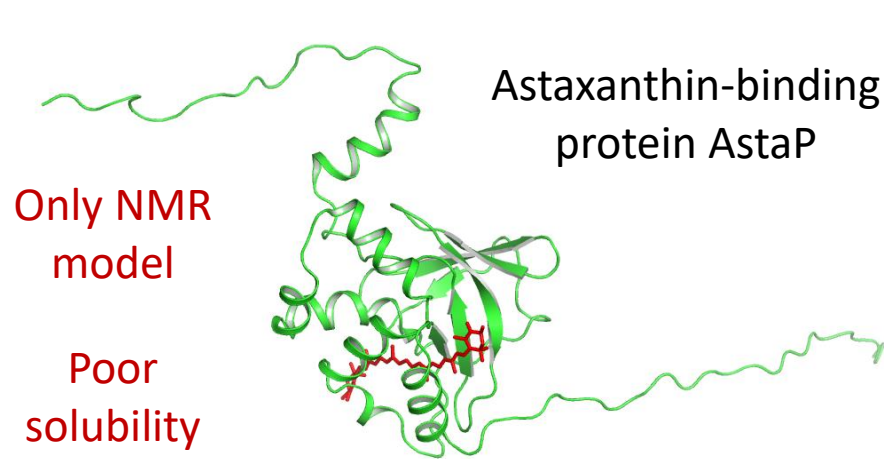
Solubilization of membrane proteins is a challenge

Finding a membrane mimic is laborious

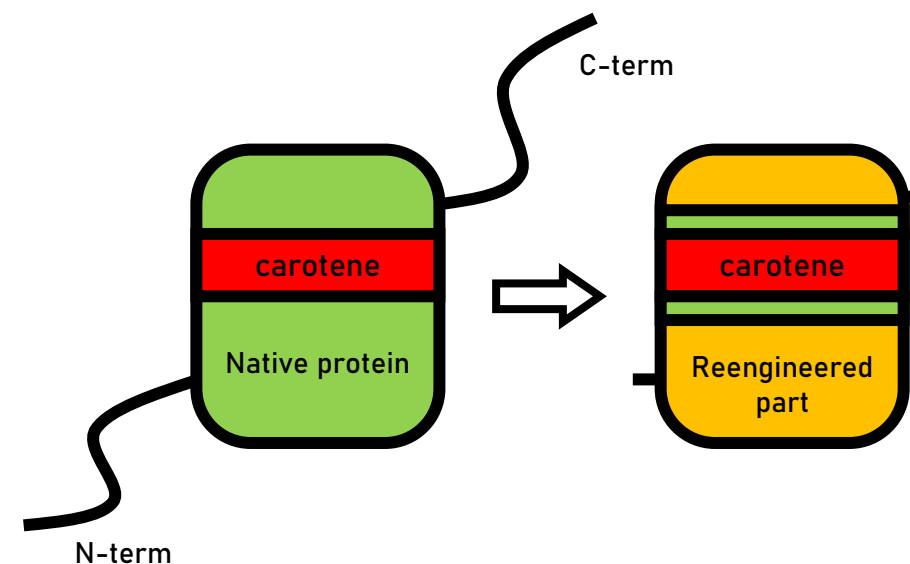
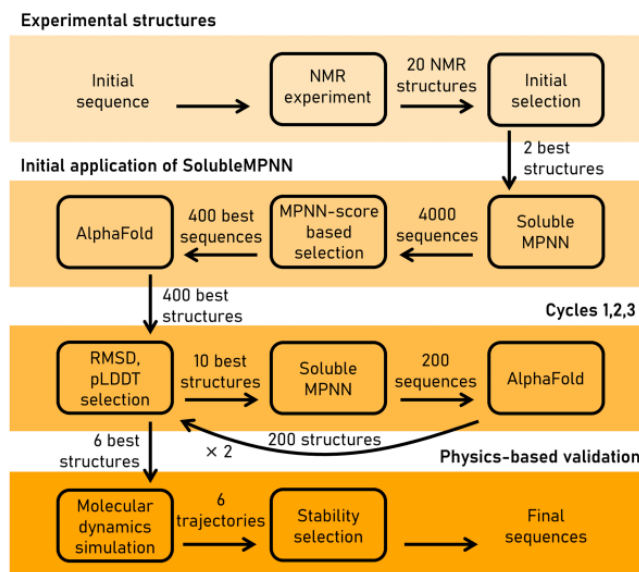
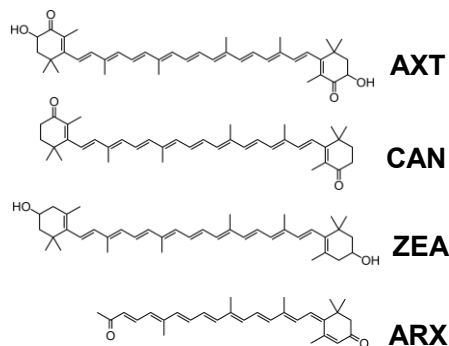
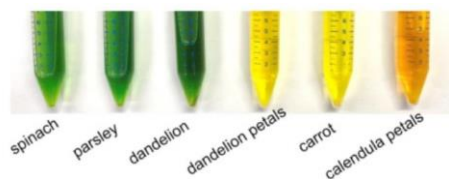
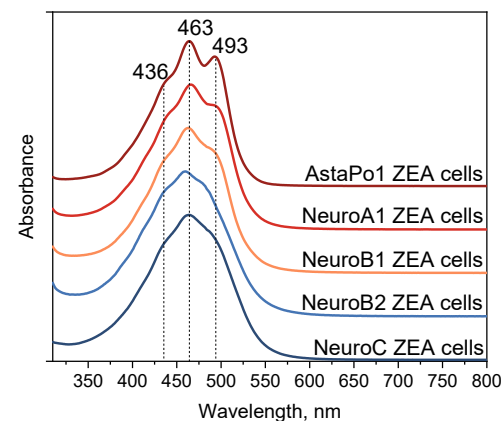
Conformations may be affected



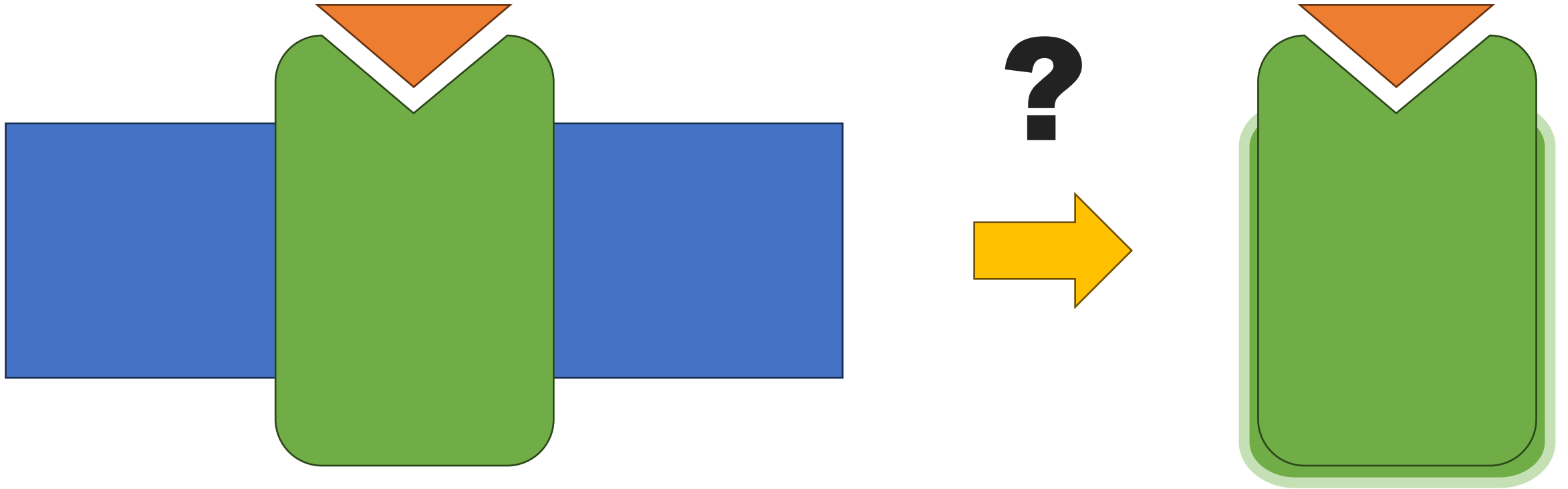
Enhancing solubility of a membrane-associated protein



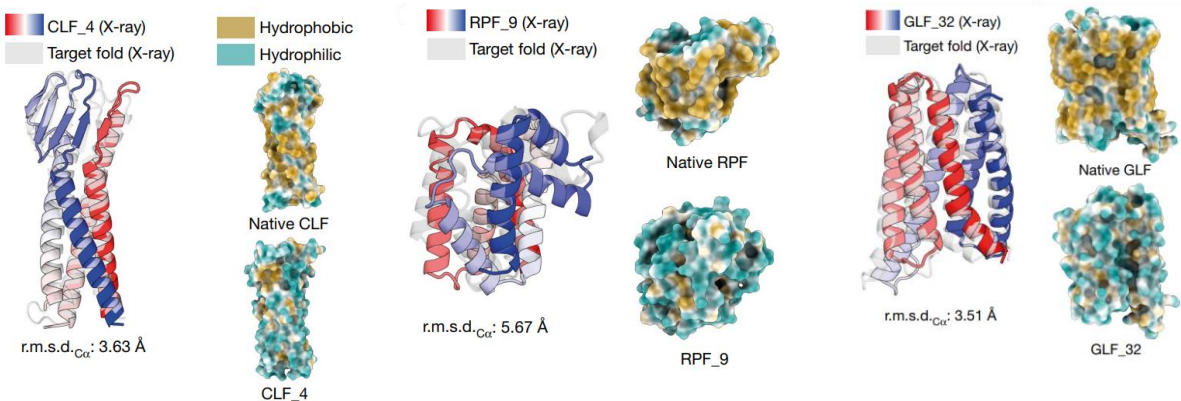
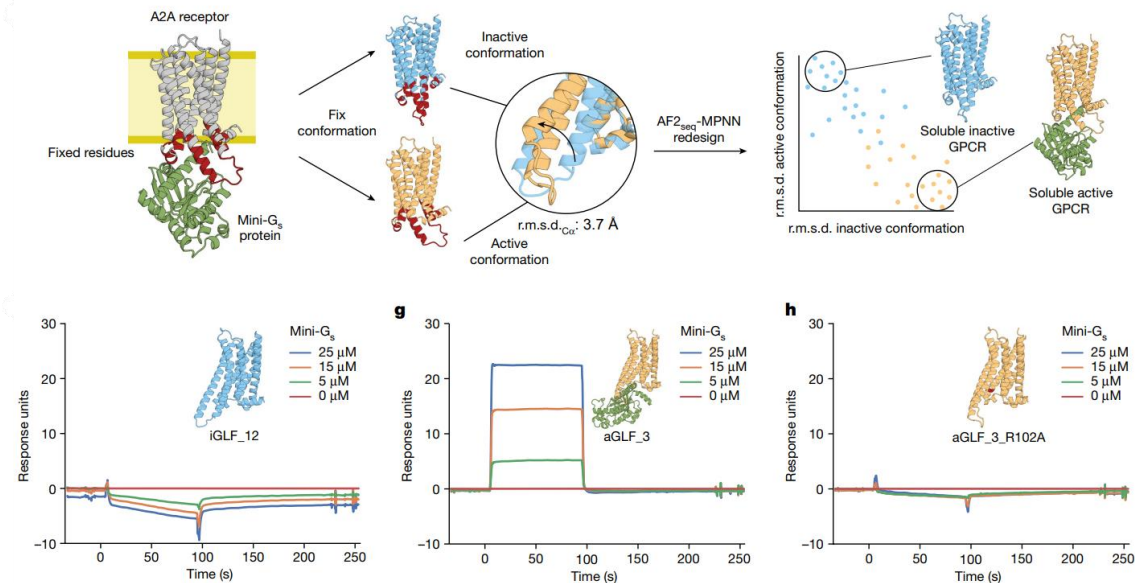
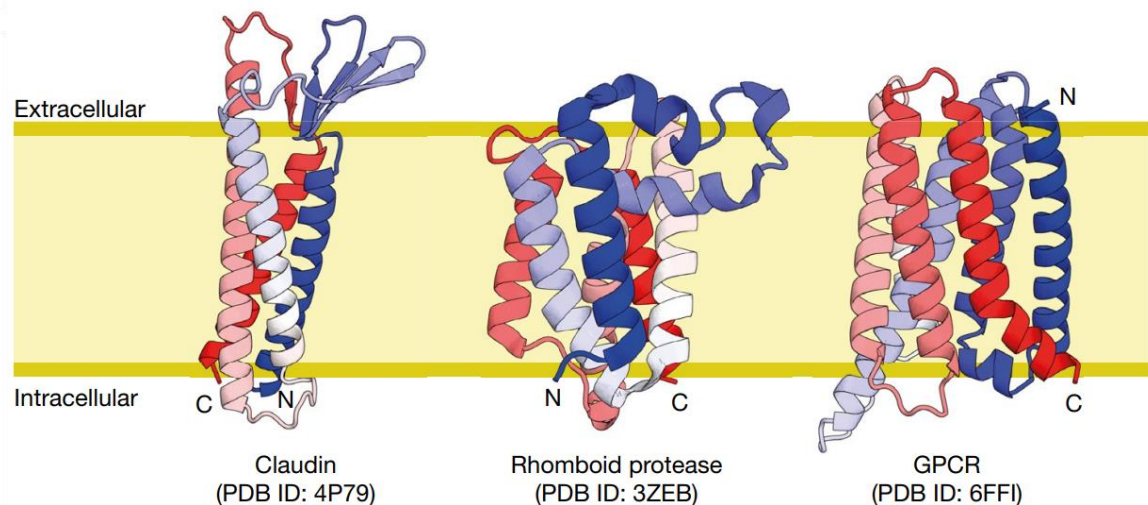
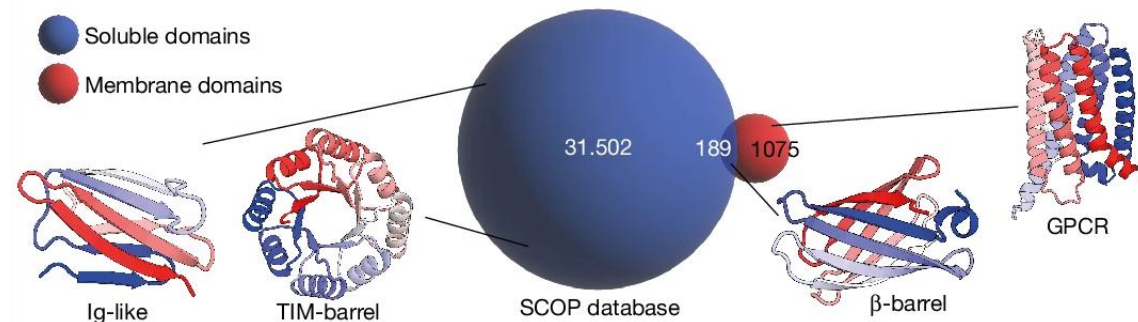
58-62% residues exchanged



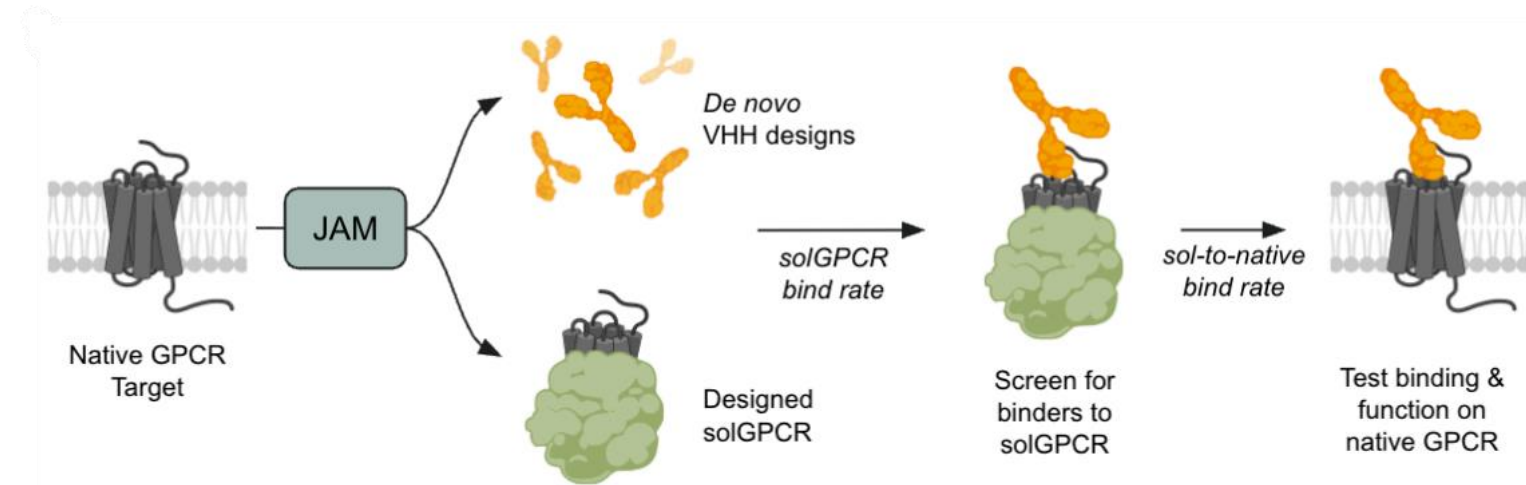
Active site mimics for challenging targets



Computational design of soluble and functional membrane protein analogues

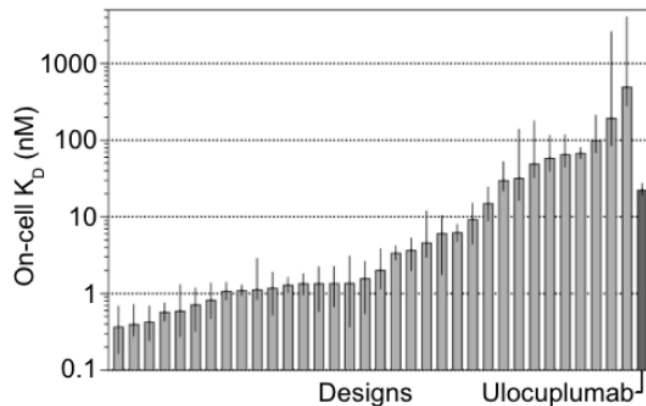


Soluble analogs for ligand screening

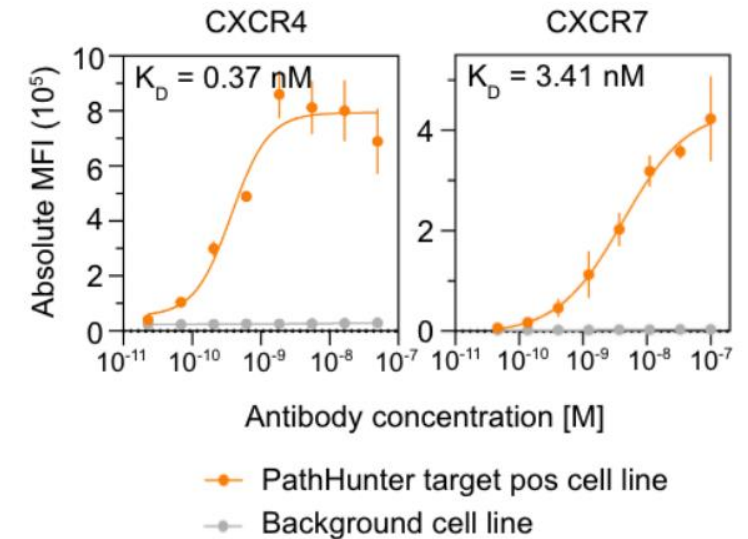
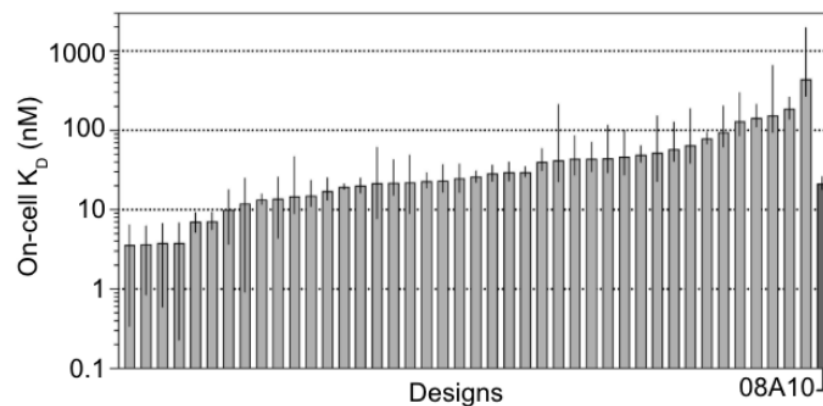


Nabla Bio

PathHunter CXCR4



PathHunter CXCR7

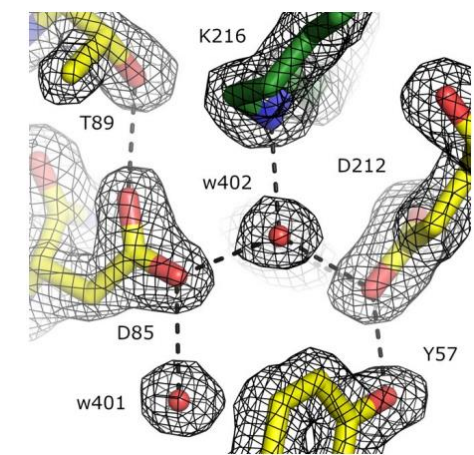
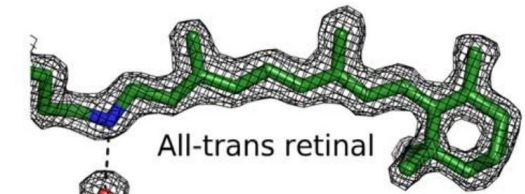
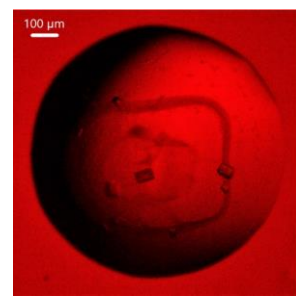
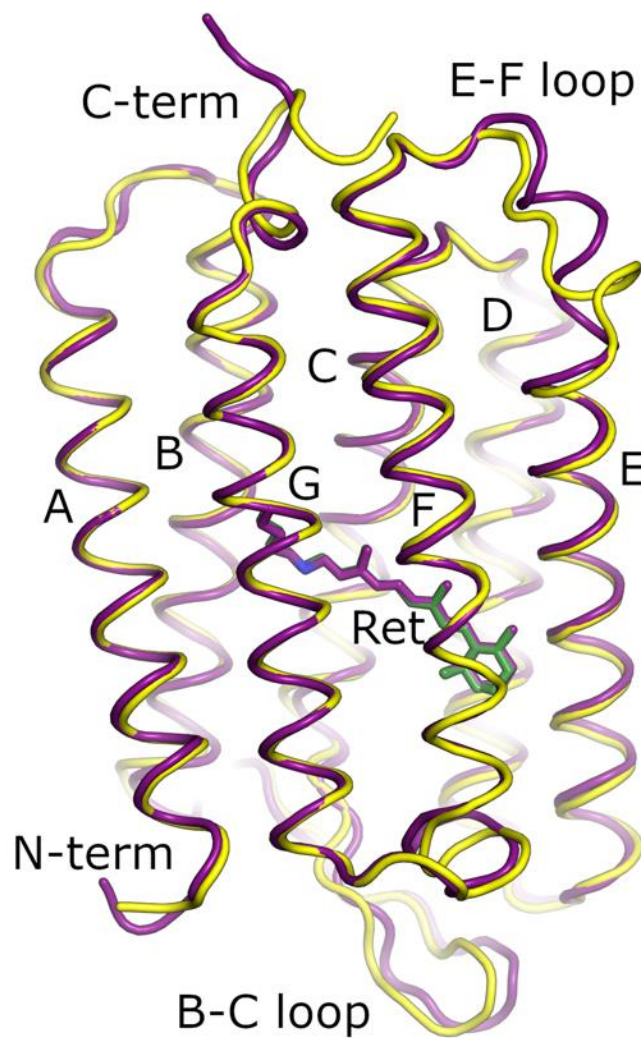
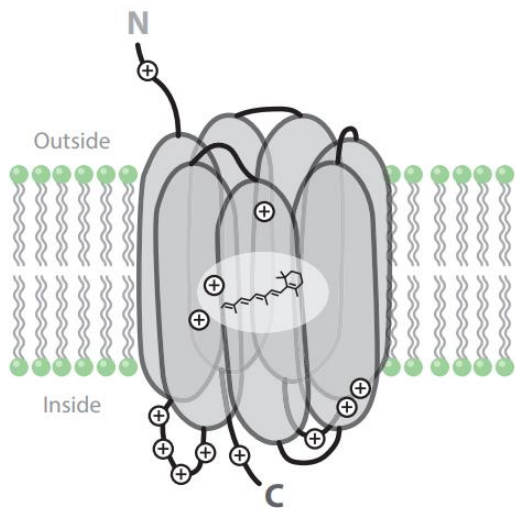
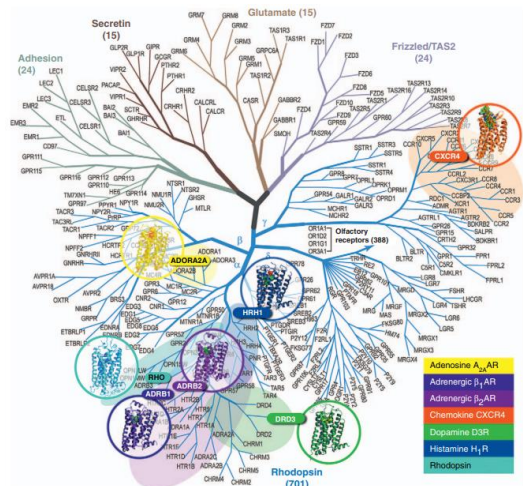


<https://doi.org/10.1101/2025.01.21.633066>

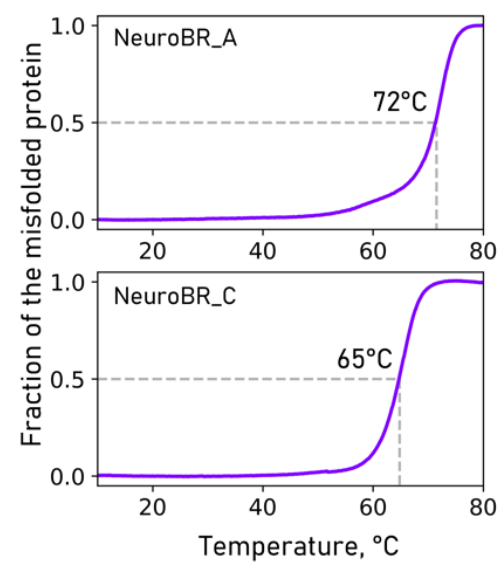
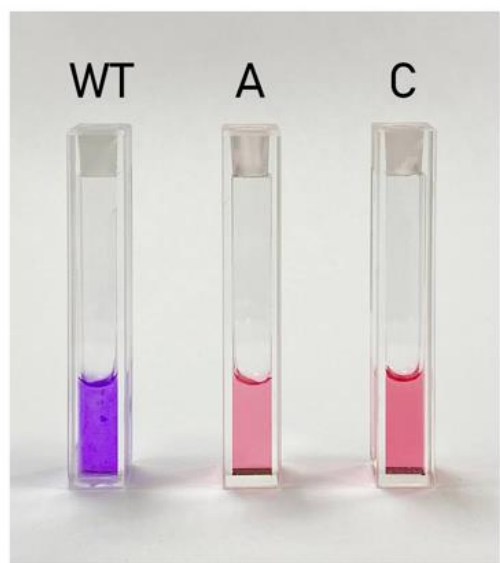
See also: <https://doi.org/10.1002/pro.70184>

<https://doi.org/10.1101/2025.08.18.670068>

Keeping intramembrane binding site during “solubilization”



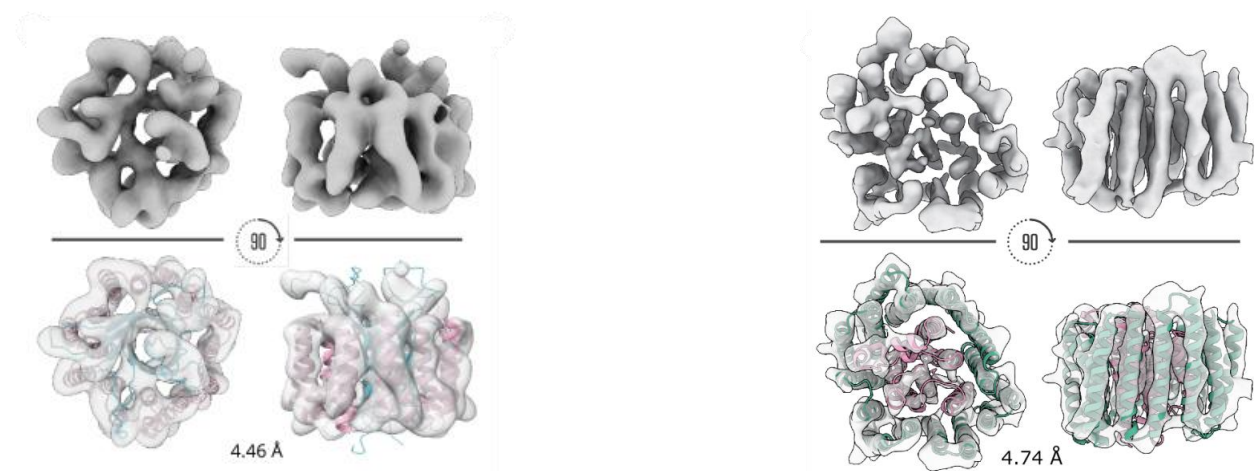
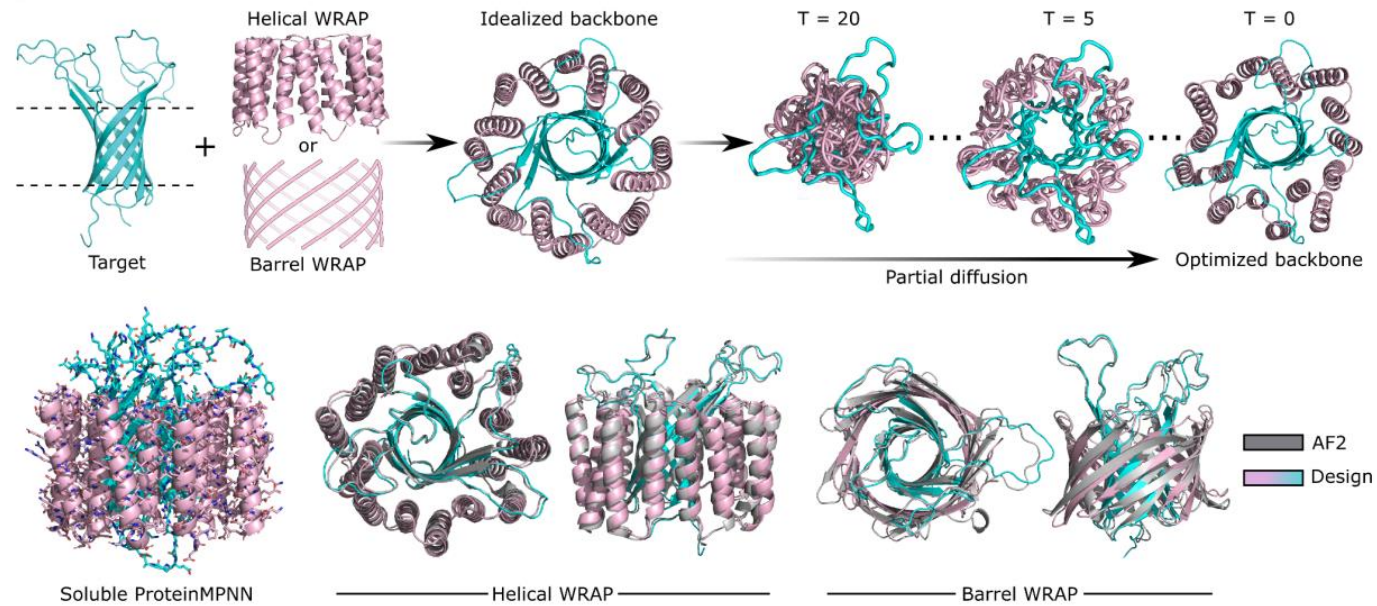
10.1146/annurev-micro-031721-020452, 10.1016/j.tips.2011.09.003



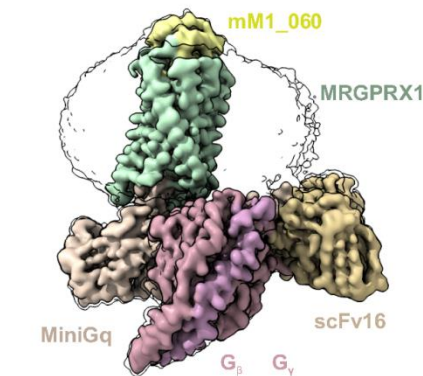
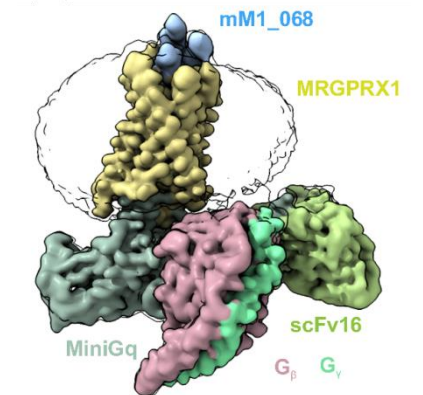
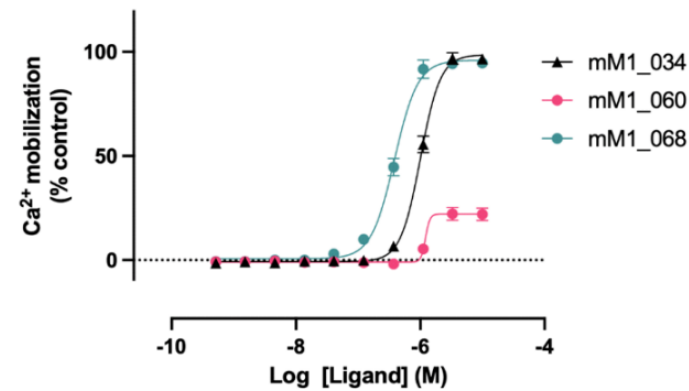
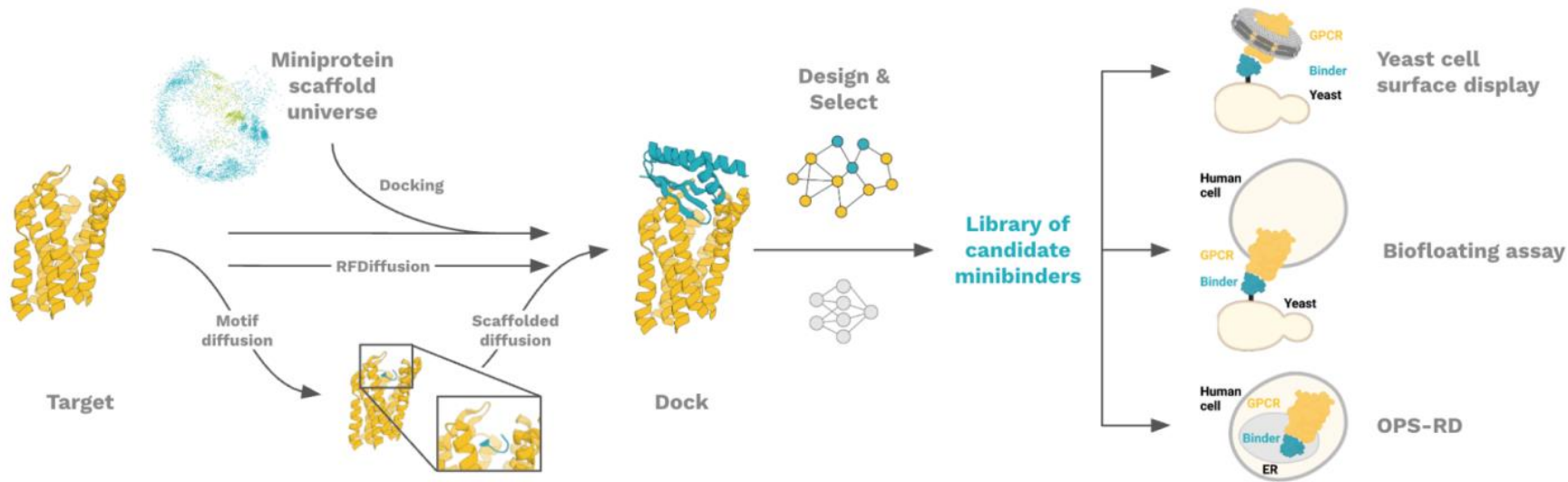
42-45% seq. id., 2/3

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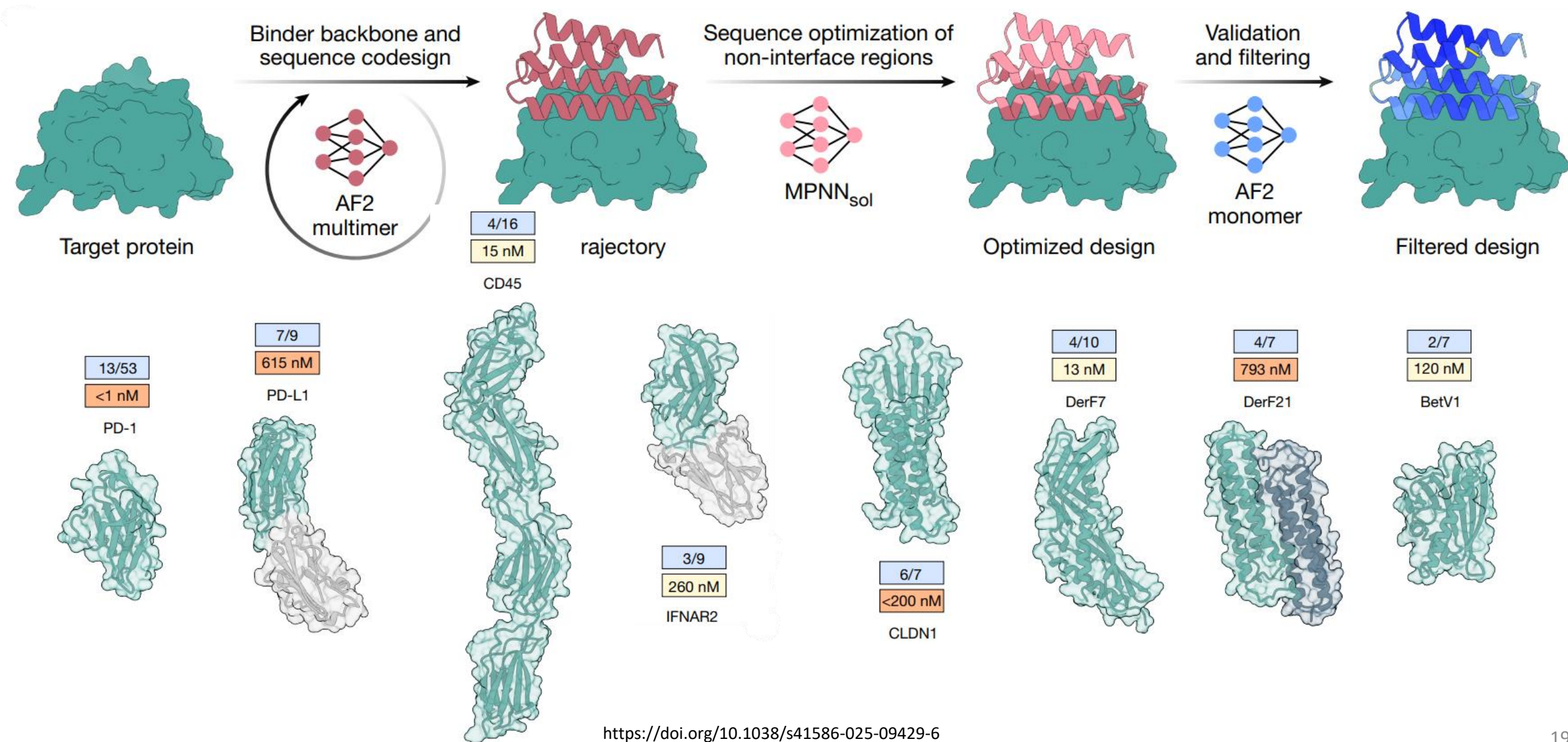
Wrapping membrane proteins for solubility



Binder proteins: GPCR activators and inhibitors

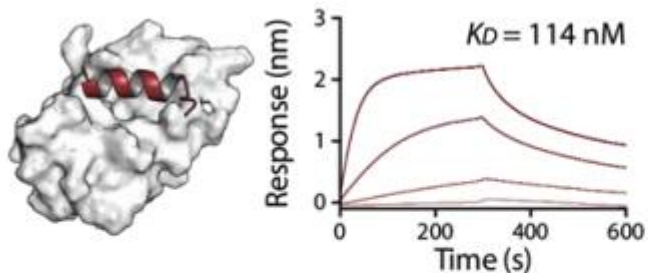


Simplified binder design with BindCraft

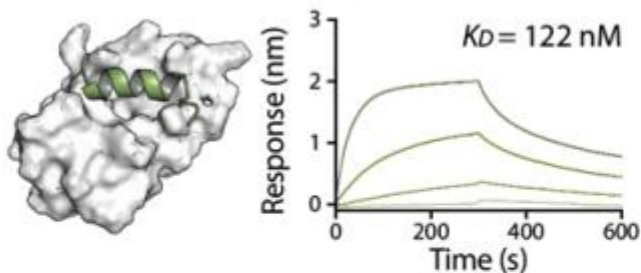


Peptide binder design with BindCraft

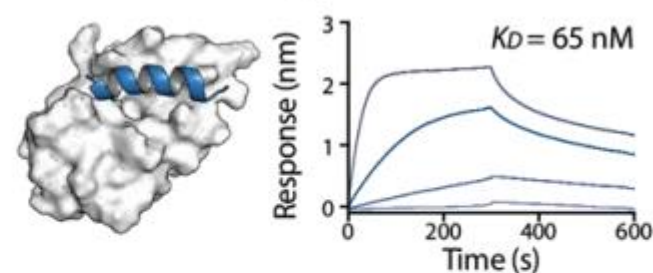
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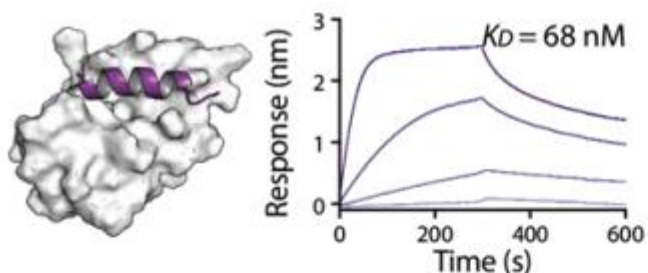
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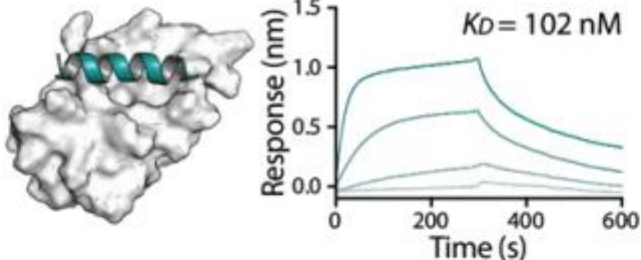
MDM2 B5 SASDEFQKEWEDLMNF



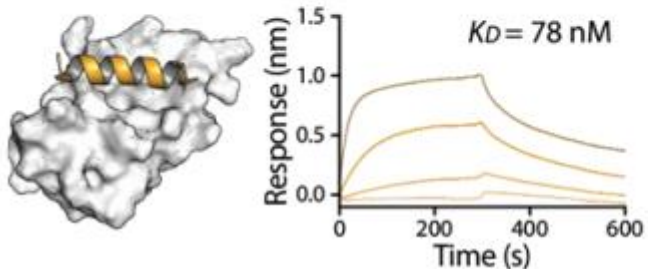
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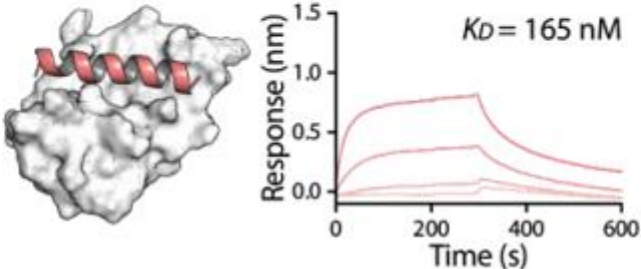
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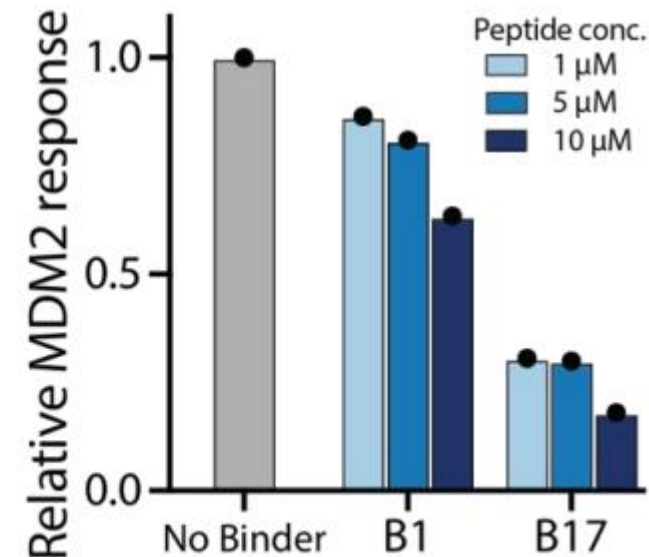
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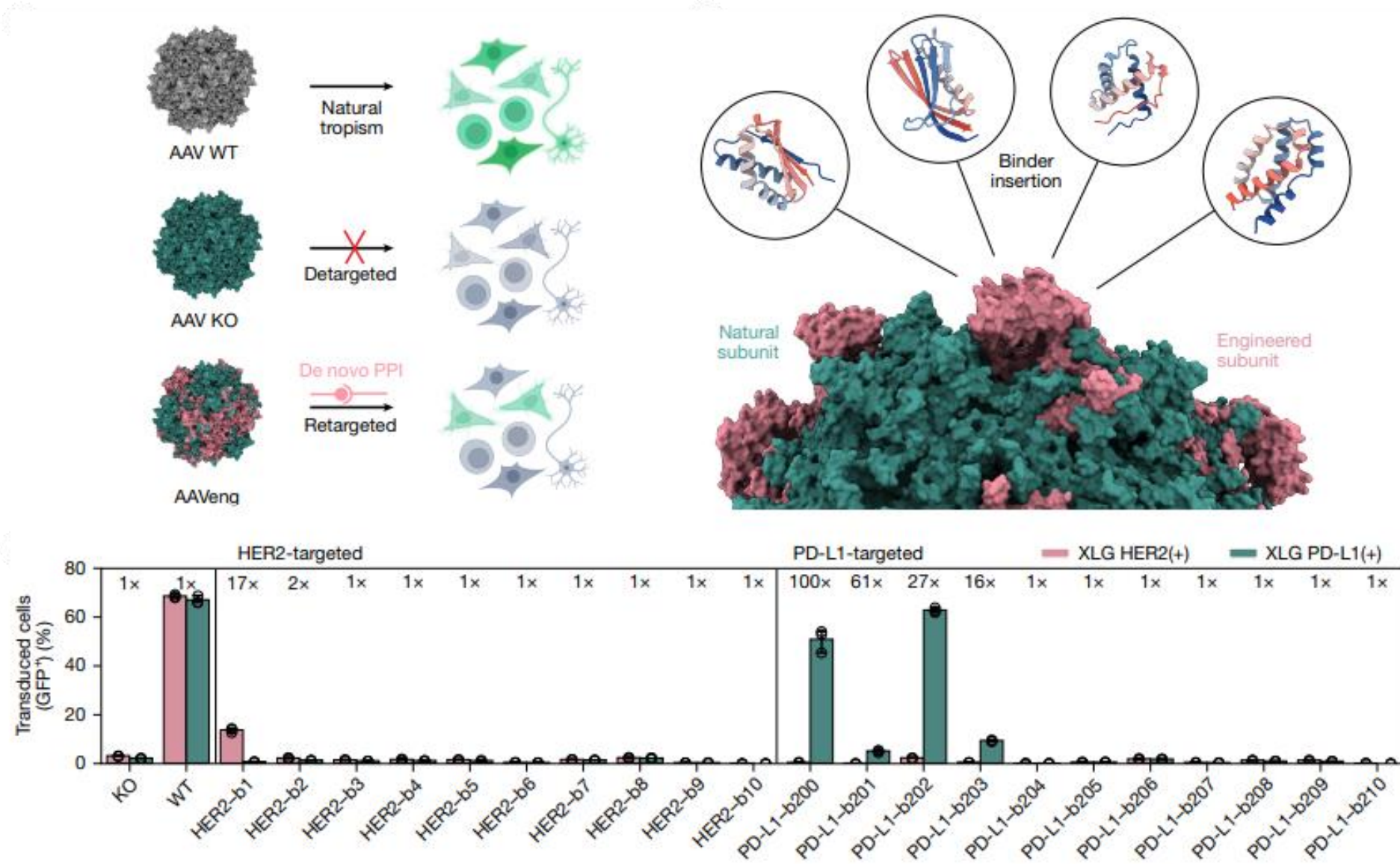
MDM2 B20 GWEGFMKQWKEFSENLEKYM



g



AAV tropism design with BindCraft



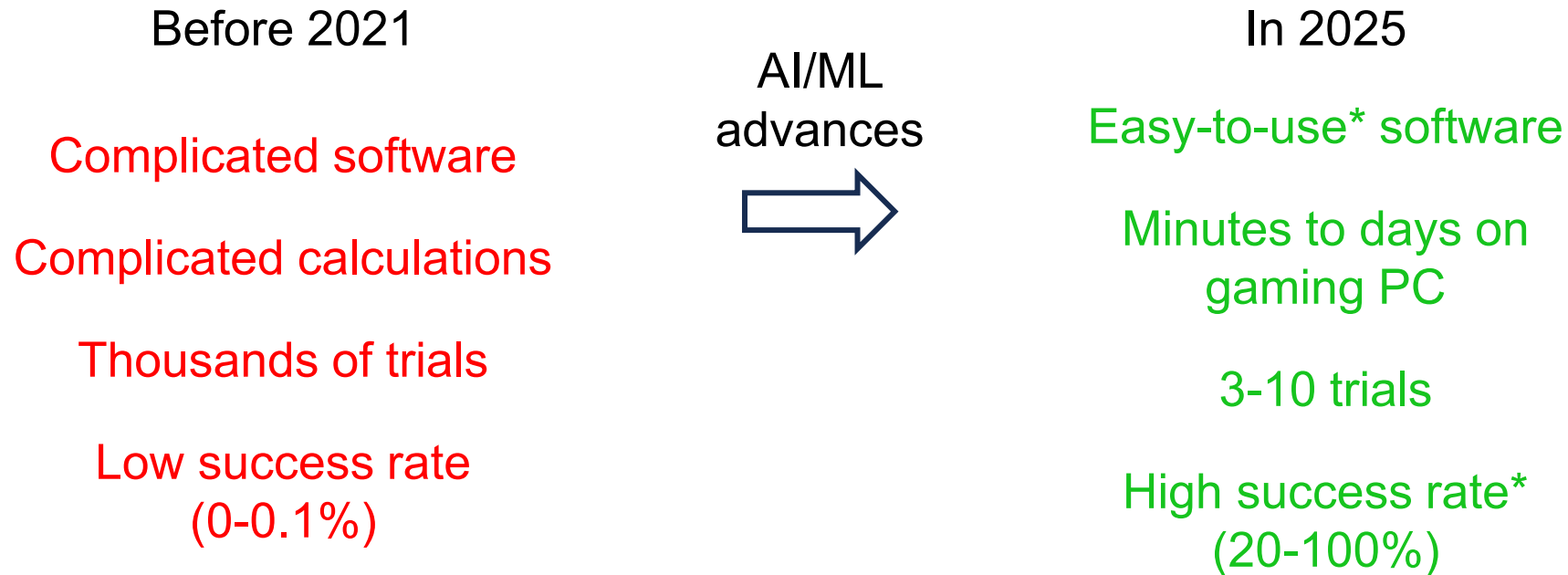
Overall success rates

Table 1. Metrics of AI-designed <i>de novo</i> binders to different targets - published					
Targets	Design method	No. designs (backbones)	Exp. Hit Rate	K_d (Top hits)	T_M (°C)
RFdiffusion, Watson et al., <i>Nature</i> (July 2023)					
Influenza An H1 Hemagglutinin (HA), IL-7R α , PD-L1, InsR, TrkA	RFdiffusion	Several thousand	7.4–34.7%	27nM-1.4 μ M (BLI)	–
Helical peptide binders, protein biosensors, Vázquez Torres, Leung, Venkatesh et al., <i>Nature</i> (December 2023)					
Bid	Hallucination	~2000	8.7%	7nM (FP)	95
Bim, Parathyroid hormone, PTH	Unconditional RFdiffusion	~2000	26.0–58.3%	<500pM (FP)	95
Glucagon, GCG, Neuropeptide Y, NPY	Parametrically designed inpainted binders (RF _{joint})	–	1.0%	231nM-3.5 μ M (FP)	–
	Partial diffusion optimization of inpainted binders	~2000	20.8–26.0%	<500pM-5.3nM (FP)	–
GCG, NPY, PTH, Peptide YY, PYY	Fine-tuned RFdiffusion (trained on 2 chain systems from PDB)	–	–	24.5nM-3.82 μ M (FP)	–
RFdiffusion All-Atom, non-protein ligands, Krishna, Wang, Ahern et al., <i>Science</i> (March 2024)					
Digoxigenin, heme, bilin	RFdiffusionAA	25,000, ~10,000, 2,776	0.07–66.7% ^a	343nM (ITC) (DIG_1)	95
TNFR agonist and antagonist binders, Glögl et al., <i>Science</i> (December 2024)					
TNFR1	RFdiffusion (hotspot residues)	30,000	6.7%	29, 24.5nM, >10 μ M (SPR)	95
	Partial diffusion optimization	25,000	29.8%	5nM, <10pM, 20nM (SPR)	95
TNFR2, OX40, 4-1BB	Partial diffusion (RFdiffusion), using TNFR1 designs as input	25,000	2.1–45.8%	198pM-44nM (SPR)	95
Elapid snake venom binders, Vázquez Torres et al., <i>Nature</i> (January 2025)					
ScNtx, α -cobratoxin (P01391)	RFdiffusion (β -edge pairing)	~2000	2.3–2.4%	842nM-1.3 μ M (BLI)	–
	Partial diffusion optimization	~2000	10.5–14.1%	0.7–6.7nM (BLI, SPR)	78- >95
Consensus cytotoxin	RFdiffusion (hotspot residues)	~2000	5.6%	271nM (SPR)	61
	CTYX with an introduced disulfide bond to reduce flexibility	NA	NA	740nM (SPR)	70.3
Macrocyclic binders, Rettie, Juergens, Adebomi et al., <i>Nature Chemical Biology</i> (June 2025)					
MCL1, MDM2, GABARAP, RbtA	RFpeptides (+/- hotspot residues)	10,000–20,000	21.4–66.7%	6nM-2 μ M (SPR)	–
Anti-microbial/heme-uptake transporter binders, Fox et al., <i>Nature Communications</i> (July 2025)					
ChuA	RFdiffusion (hotspot residues)	25,000	8.3%	64.4–155nM (BLI)	–

Table 2. Metrics of AI-designed <i>de novo</i> binders to different targets - preprints					
Targets	Design method	No. designs (backbones)	Exp. Hit Rate	K_d (Top hits)	T_M (°C)
Binders to intrinsically disordered proteins and regions, Liu, Wu, Choi et al., <i>bioRxiv</i> (July 2024)					
Amylin (hIAPP), C-peptide, VP48, G3bp1, IL2RG, Prion	RFdiffusion (either with varied constraints or unconstrained)	~10,000–50,000	1.0–5.1%	14nM-16 μ M (BLI)	95
	Two-sided partial diffusion optimization	~10,000–50,000	3.2–61.5%	3.8nM-2 μ M	95
AlphaProteo (closed-source), Zambaldi, La, Chu, Patani, Danson, Kwan, Frerix, Schneider, Saxton, Thillaisundaram, Wu et al., <i>arXiv</i> (September 2024)					
BHRF1, SARS-CoV-2 spike RBD, IL-7R α , PD-L1, TrkA, IL-17A, VEGF-A, TNF α	AlphaProteo (hotspot residues)	–	0-88%	82pM-26nM (HTRF)	95
BindCraft, Pacesa, Nickel, Schellhaas et al., <i>bioRxiv</i> (October 2024)					
PD-1, PD-L1, IFNAR2, CD45, BBF-14, CrSAS-6, Der f7, Der f21, Bet v1, SpCas9	BindCraft (+/- hotspot residues)	300–6000	10-100%	<1nM-5.7 μ M (SPR)	90
Improved binder design with beta-pairing RFdiffusion, polar targets, Sappington et al., <i>bioRxiv</i> (October 2024)					
KIT (SCFR, CD117), PDGFR α , ALK-2, ALK-3, FCRL5, NRP1, α -cobratoxin	RFdiffusion (β -strand pairing, target residues)	~10,000	–	76pM-193nM (SPR)	63.2- >95
Binders for use in CAR-T therapies, Mergen, Abele et al., <i>bioRxiv</i> (November 2024)					
BCMA (CD269)	RFdiffusion (Google Colab, hotspot residues)	1-28 designs for 18 binders	16.7%	–	–
pMHC-I binders, CAR-T therapies, Johansen, Wolff, Scapolo, Fernández-Quintero et al., <i>bioRxiv</i> (December 2024)					
Cancer/Testis Antigen 1B NY-ESO-1 ⁽¹⁵⁷⁻¹⁶⁵⁾ /HLA-A*02:01, Metastatic melanoma neoantigen RVTDESILSY /HLA-A*01:01	RFdiffusion (against non-achor peptide residues)	2,100, 5,500	2.1–2.3%	–	–
Anti-CRISPR binders (Alcra), Taveneau et al., <i>bioRxiv</i> (December 2024)					
LbuCas13a	RFdiffusion (hotspot residues)	10,000	5.2–10.4%	–	70
TCR-mimic binders, Householder et al., <i>bioRxiv</i> (December 2024)					
NY-ESO-1 ⁽¹⁵⁷⁻¹⁶⁵⁾ /HLA-A*02	RFdiffusion (hotspots residues)	100	40%	9.5nM (SPR)	–
GPCR agonist and antagonist binders, Muratspahić, Feldman, Kim, Qu, Bratovianu, Rivera-Sánchez et al., <i>bioRxiv</i> (March 2025)					
MRGPRX1, CXCR4, GLP1R, GIPR, GCGR, CGRPR, CGRPR	RFdiffusion (scaffold-guided)	26,000–100,000	0.008–2.1% ^a	5.3–27nM (SPR)	95
	Partial diffusion optimization	3,000	46.2%	–	–
Chai-2 (closed-source), Chai Discovery Team et al., <i>bioRxiv</i> (July 2025)					
PDGFR, IL-7R α , PD-L1, InsulinR, TNF α	Chai-2	–	22-95%	18.3pM-11.7nM ^b (BLI)	–

Gene synthesis cost as low as 1-3 k\$ per screening round

Progress in protein engineering



Summary

- 1. Protein design strongly advances drug discovery efforts and provides new avenues (stabilization, mimics, binders)**
- 2. Rapid development of new methods leads to higher competition and search for new biological insights**
- 3. Protein engineering gets simple: try it yourself!**

Acknowledgements



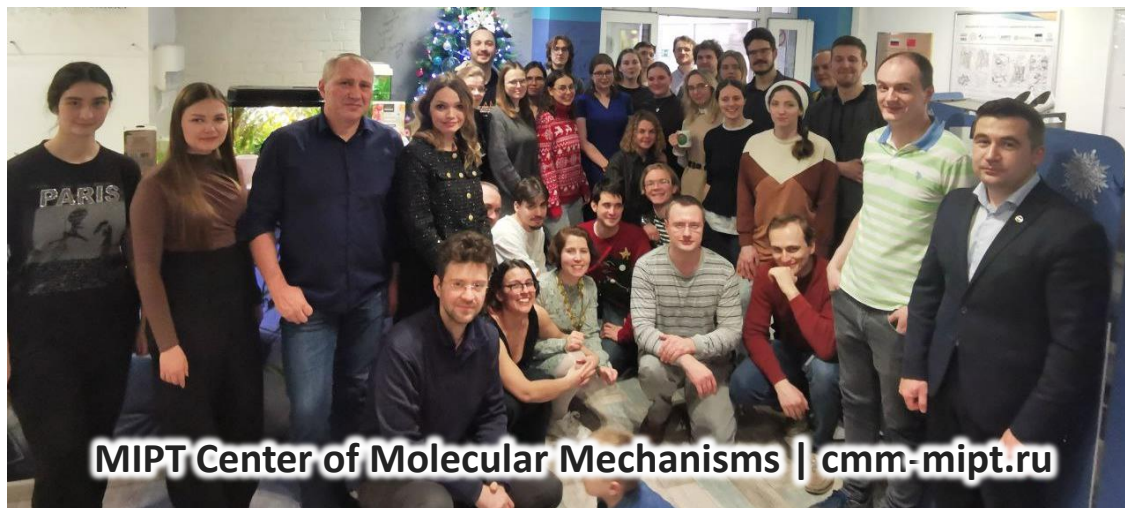
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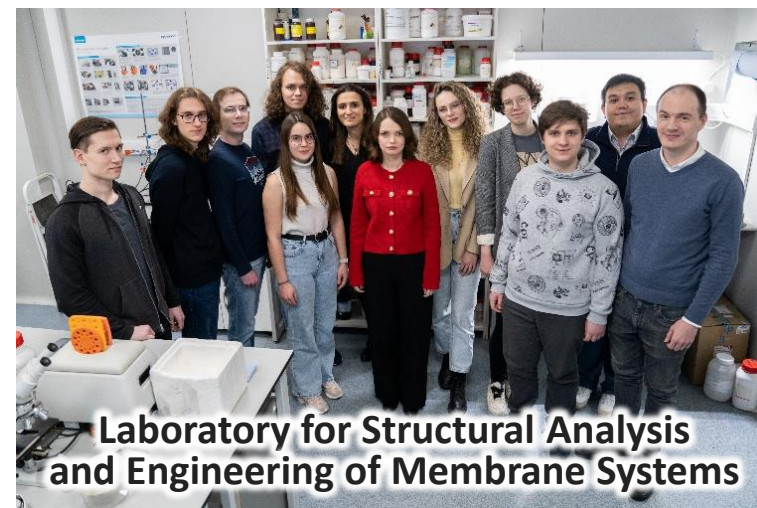
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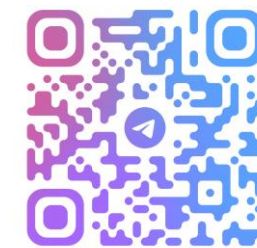
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